

## Understanding the role of Molecular and Genetic Alterations in Proliferation and Progression of Oral Squamous Cell Carcinoma : A review article.

### Abstract:

Oral carcinogenesis is known as multifactorial process which engaged plentiful genetic events that transform normal activity of tumor suppressor genes and oncogenes. It is observed that aggregation of genetic alterations is the ground for advancement of a normal cell to cancer cells, which is known as a multi-step carcinogenesis. Because of this event, growth factor production increases as well as increase in total of receptors on cell surface and increased intracellular signal messengers. The present review scrutinize the existing documentation in the literature related to the oral squamous cell carcinoma. English language articles were searched in various databases such as Pubmed, Scopus, Science direct and Google scholar. The keyword used for searching are "oral squamous cell carcinoma", "Genetics and Oral squamous cell carcinoma", "Molecular mechanism in oral squamous cell carcinoma". The present review spotlights on understanding the molecular mechanism and the genetic factors which is responsible for alteration in the cell which leads to oral squamous cell carcinoma.

**Key-words:** Oral squamous cell carcinoma, Genetic alterations, Molecular mechanism, Proliferation, Signaling

### Introduction:

The squamous cell carcinoma is the most ubiquitous malignancy seen in the head and neck, pharynx, and oral cavity. Generally, 3 lakh new oral cancer cases, as well as roughly 68 thousand loss of life worldwide are predicted annually[1]. Squamous cell carcinoma is defined as "a malignant epithelial neoplasm exhibiting squamous differentiation as characterized by the formation of keratin and/or the presence of intercellular bridges"[2].

In modern years, a considerable breakthrough has been made in recognition of the genetic and molecular basis of the advancement of oral squamous cell carcinoma. According to multi-step carcinogenesis, aggregation of genetic alterations in the ground for growth from a normal cell to a cancer cell[3]. Progression or advancement is facilitated by more and more abnormal functions of genes that positively as well as negatively balanced the condition of proliferation, apoptosis, angiogenesis, genome stability, invasion, and metastasis.

There are various ways by which gene function can be transformed like oncogenes can be activated by amplification or mutation and tumor suppressor genes may be inactivated by deletion, mutation, or methylation[4,5,6].

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Universal alterations for oral cancer include TP53 inactivation, the progress of chromosomal material at 3q26 as well as 11q13, and losses at 13q21, 3p21, and 14q32[7]. The tumor microenvironment is considered an essential contributor to fostering proliferation, survival, neoplastic process, and migration[8].

Recent experiments have developed the perception that tumor microenvironments are the important components of the progression of tumors along with inflammation and hypoxia. Plentiful cancers arise from the location of inflammation, chronic irritation, and infection. Some tumors also consist of hypoxic microenvironments which are related to poor prognosis as well as resistance to treatment[9,10]. Hence, oral carcinogenesis is an eminently complex multifactorial process that occurs when several genetic alterations affect the epithelial cells.

### **Molecular Genetics/genetic Alterations**

#### **P16/cyclin D1/prb/p53**

p16 and p53 are two crucial tumor suppressor proteins that are intermittently inactivated in head and neck cancer. p16 is encoded by the CDKN2A gene which is positioned on chromosome 9p21. p16 is implicated in cell growth as well as cell cycle control. Individually p16 blocked the growth from G1 to S phase of the cell cycle by inhibiting a separate protein known as Cyclin D1. Hence, interruption of activity of p16 outcomes in loss of cell senescence afterward leads to dysplasia[11,12].

The p53 is a tumor suppressor gene having a molecular weight of 53kDa. The p53 gene encodes a protein that seized the cell cycle at the late G1 phase in those cells which have sub-lethal impairment in their genome as far as their entire repair or it induces apoptosis in unrecoverable injury cases[13,14]. Mutations of the p53 gene are scattered over a considerable hundred base pairs in the mid-region of the gene. They occur especially in exons 5-8, which are recognized as hot regions. During metastasis, these mutations remain steady. Mutation of the p53 gene appears during the early phase or stages in the progression of head and neck squamous cell carcinoma for the reason that they are earlier present in premalignant lesions[15,16].

Cyclin D1 protein is encoded by gene CCND1, whose activity is constrained by p16. It is observed that the CCND1 gene is amplified in approximately 25-43% of oral squamous cell carcinoma<sup>17,18</sup>. Cyclin D1, which is the product of the CCND1

gene, was seen to be overexpressed during the early phase of oral carcinogenesis. Also, Cyclin D1 is found to be associated with tumorigenic proliferation[19,20]. It was observed that expression of cytoplasmic Cyclin D1 was increased in oral squamous cell carcinoma with advanced stages, increased mitosis, poor differentiation, and invasive cell morphology, implying that increased level of cytoplasmic Cyclin D1 encouraged cell migration and invasion. Furthermore, in some studies, Cyclin D1 expression and CCND1 amplification were associated with poor prognosis and decreased rate of survival[21,22].

#### **Notch:**

Notch signaling is the eminently conserved pathway of connection amidst neighboring cells and regulating cell proliferation and fate[23,24]. In some cancers, especially human T cell acute lymphoblastic leukemia, the activated mutation in NOTCH1 is seen[24]. NOTCH1 may act as a tumor suppressor as inactivated mutations in NOTCH1 are seen in 11-19% of cases having Oral squamous cell carcinoma[25,26].

NOTCH1 is expressed in the basal cells of the oral squamous epithelium. It is important to note that the NOTCH1 expression is inhibited in oral epithelial dysplasia as well as in oral cancer[27]. Pickering had observed that 9% of patients having oral squamous cell carcinoma suppresses inactivating mutations in NOTCH1. In vitro proliferation of oral squamous cell carcinoma was inhibited by functional NOTCH1 signaling. NOTCH1 mutations were infrequent in Singaporean patients having oral tongue squamous cell carcinoma, whereas increased rates of NOTCH1 mutation is seen in Chinese patient having oral squamous cell carcinoma which advocates that NOTCH1 inactivation have composite genetic interactions in the occurrence of oral squamous cell carcinoma[28,29].

In oral squamous cell carcinoma, overexpression of Nrf2 was seen to develop a cancer phenotype. Hence, tumorigenesis may emerge from numerous forms of dysregulation of the Notch signaling pathway that suggest its concerned regulation is important for proper cell function[30].

#### **Wnt/ $\beta$ -catenin:**

The signaling pathway in Wnt/ $\beta$ -catenin is a conserved pathway that coordinates cell fate determination, differentiation, and proliferation. Upregulation of signaling pathways leads to oncogenesis in oral squamous cell

carcinoma.  $\beta$ -catenin is encoded by CTNNB1. It is found that mutations in CTNNB1 are rare in oral squamous cell carcinoma[26].

### **Epidermal growth factor receptor:**

The epidermal growth factor receptor is a tyrosine kinase receptor. When activated, it upregulates considerable downstream signaling pathways that include AKT, MAPK, Jak/STAT, and ERK which are necessary for tumor growth, apoptosis, angiogenesis, proliferation, survival, invasion, and metastasis[31,32,33]. In 90% of cases of oral squamous cell carcinoma, EGFR levels are raised up[34]. Poor survival of patients and tumor aggressiveness are seen in the high expression of EGFR[35].

### **PI3K/AKT/mTOR:**

The PI3K/AKT/mTOR pathway is a well-expressed signaling axis that is involved in the growth, survival, proliferation, and drug resistance of different cancer types[36,37]. In this pathway, activation of the PI3 family of protein kinase leads to PIP3 formation. This formation, in turn, activate AKT which further results in the activation of the different downstream target along with mTOR (mammalian target of rapamycin)[38]. Some studies have demonstrated that there is a direct role of activation of AKT signaling in oral squamous cell carcinoma. In immunohistochemistry analysis, staining of AKT, pmTOR, and pAKT are seen in considerable count in oral squamous cell carcinoma[39].

Via activation of AKT signaling, numerous tumor-associated pathways are shown to promote survival, invasion, growth, or drug resistance in oral squamous cell carcinoma[40,41]. Recently identified contributors which activates AKT signaling are FoxM1, RACK1, ZNF703, Nox1, Muc1, PDGF-D, CCL18[42,43,44].

### **p33ING1b gene:**

ING tumor-suppressor proteins(ING 1-5), which is a growth inhibitor protein, have been discovered in past decades. It was suggested that ING1 was a negative growth regulator which acts as a class II tumor suppressor. It directly play role in apoptosis, oncogenesis, cell cycle regulation, and DNA repair. The p33ING1b protein is a widely expressed isoform of ING1 in human normal tissues. The ING1 proteins were intermittently downregulated. ING1 proteins were elevated in papillary thyroid carcinoma, melanoma, and ductal breast carcinoma[45,46]. ING1 was less periodically mutated in various human malignancies, such as colon carcinoma, brain

tumor, oral squamous cell carcinoma, gastric tumor, and neuroblastomas[47,48]. ING1 gene is located on human chromosome 13q33-34. It is observed that this region is involved in the advancement of various tumors[49,50].

### **Cylindromatosis (CYLD) gene:**

CYLD (cylindromatosis) gene is a known tumor suppressor gene[51]. It inhibits the activation of NF- $\kappa$ B. The NF- $\kappa$ B plays important role in tumorigenesis, immune response, inflammation, and protection against apoptosis[52,53,54].

### **Neural cell adhesion molecule (NCAM)**

NCAM or neural cell adhesion molecule is a family of cell surface glycoproteins. NCAM plays a key role in nervous system development, axonal outgrowth, fasciculation, regulating cell migration, and branching[55,56]. The expression of NCAM is upregulated by TGF- $\beta$  (transforming growth factor)[ 57]. It has been observed that NCAM expression is seen in numerous human neoplasms. Also, NCAM is participating in perineural invasions in numerous neoplasms such as gallbladder cancer, bile duct cancer, adenoid cystic carcinoma[58,59,60]. NCAM mediates the cell's adhesion through a Ca<sup>2+</sup>-independent hemophilic (NCAM—NCAM) binding mechanism. Also, NCAM mediates the adhesion between neurons and extracellular matrix through heterophilic binding[55].

### **Receptor-binding cancer antigen expressed on SiSo cells (RCAS1):**

RCAS1 is known as type II membrane protein which is isolated as a human tumor-associated antigen. RCAS1 act as a ligand for a receptor that is present on immune cells such as B, T, and NK cells. The growth of receptor-expressing cells is inhibited by RCAS1[61,62]. It is observed that RCAS1 was regularly expressed in oral squamous cell carcinoma[63].

### **Tumor necrosis factor-related apoptosis inducing ligand (TRAIL):**

TRAIL is also known as APO2 ligand or APO2L. It is a unique member of the TNF cytokine family. TRAIL has the capability to induce apoptosis. Few receptors help in the binding of TRAIL[64,65].

These are death receptor-4 (DR4), DR/5KILLER, decoy receptor-1(DcR1), DcR2. Death receptor-4 (DR4), as well as DR5/KILLER, consist of signal apoptosis and cytoplasmic death domains. DcR2 does not transducer death signal. DcR1

does not have a cytoplasmic tail and prohibits TRAIL function[66,67].

### **Interleukin (IL)-12 and IL-23:**

The relationship between innate immunity, chronic inflammation, and cancer is now universally accepted. Various cancers originate at the chronic inflammation location[68,69]. It is observed that persistent use of anti-inflammatory drugs lessens the frequency of human tumors<sup>70</sup>. IL-23 has been recognized as an important factor that links tumor-associated inflammation and a loss of tumor immune surveillance. IL-23 consists of a p19 subunit which is associated with the IL-12p40 subunit. IL-12 is an association of IL-12p35 and IL-12p40 subunit[71]. The various signal-transduction components used by IL-23 include Tyk2, Janus kinase2, and Signal transducer as well as an activator of transcription Stat1, Stat3, Stat4, and Stat5[72]. In some studies, it has been observed that IL-12 has dominant antitumor activity in various murine tumor models which leads to regression of established tumors. Also, it inhibits the development of experimental and spontaneous metastasis[73,74].

### **Nuclear factor (NF)-kappaB:**

Nuclear factor-kB (NF-kB) plays important role in immune response, tumorigenesis, inflammation and atability against apoptosis[75,76,77].

### **Hypoxia inducible factor (HIF)-1alpha:**

Hypoxia inducible factor (HIF)-1alpha plays a pivotal role in biological processes specifically carcinogenesis and angiogenesis[78,79]. HIF-1a upregulates various essential factors for tumor expansion which includes VEGF. In a few cancers, tumor aggressiveness is seen to be associated with overexpression of HIF-1a[80,81,82].

### **Epigenetics:**

Epigenetic alterations to the genome bring one of the considerable mechanisms for gene expression regulation. Various processes like methylation and demethylation of gene promoter province play an important role in gene silencing or expression of the gene. This gene silencing as a result of methylation is one of the hallmarks of carcinogenesis[83,84].

It is observed that many times p16 is inactivated because of promoter hypermethylation in oral squamous cell carcinoma[85,86,87]. The methylation of the p16 promoter

occurs due to upregulated expression of DNA methyltransferases as a consequence of regular use of tobacco[88,89]. A large number of alternative genes are also hypermethylated in promoter regions in oral squamous cell carcinoma which includes RASSF1A, RASSF2A, CDH1, p14,p15, MLH1, and DAPK. These genes play a crucial role in different cancer-promoting signaling networks like Wnt signaling, DNA repair, Cell adhesion, and Cell cycle arrest and apoptosis[90,91,92,93].

### **Metastasis/invasion:**

Metastasis is known as a complicated process in which selected malignant cells acquire the potential to remain alive in a distant environment. The main steps in metastasis include angiogenesis and epithelial-mesenchymal transition. Angiogenesis is a process that concentrates around the activation of VEGF (vascular endothelial growth factor). VEGFA is a useful prognostic indicator for oral tongue squamous cell carcinoma. Also, overexpression of VEGFA shows poor survival[94,95].

Epithelial-mesenchymal transition characteristically involves the reduction of molecules that influence cell adhesion such as upregulation of cytoskeletal protein (Vimentin) and upregulation of mesenchymal protein (N-cadherin).

In various immunohistochemical studies, oral cancers show increased expression of Vimentin and decreased expression of E-cadherin. Hence, it recommends that cancers take advantage of epithelial-mesenchymal transition for invading and metastasizing[96,97]. Epithelial-mesenchymal transition is also eminently driven by the expression of TGF- $\beta$  i.e. transforming growth factor. TGF- $\beta$  activates cell spreading and also causes partition of cell borders. It has been observed that overexpression of TGF- $\beta$ 1 causes migration and invasiveness of oral squamous cell carcinoma and advances the malignancy[98,99]. Phosphorylation of  $\beta$ -catenin results in dissociation from the complex with E-cadherin, which leads to loss of cell adhesion and consequential invasion of tumor cell[100,101].

### **Conclusion:**

Our knowledge of molecular basis of oral squamous cell carcinoma grows promptly in past years. Various genetic proceeding that conclude in carcinogenesis comprises of oncogenes activation and deactivation of tumor suppressor genes. Recent progress in microarray technologies and DNA

sequencing provides a exceptional favourable circumstances for thousand of genes to be managed at the same time.

### References:

1. Enwonwu CO, Phillips RS, Ibrahim CD, Danfillo IS (2004) Nutrition and oral health in Africa. *Int Dent J* 54:344–351
2. Epstein JB, Zhang L, Rosin M (2002) Advances in the diagnosis of oral premalignant and malignant lesions. *J Can Dent Assoc* 68(10):617–621
3. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100:57–70.
4. Califano J, van der Riet P, Westra W, Nawroz H, Clayman G, Piantadosi G, et al. Genetic progression model for head and neck cancer: implications for field cancerization. *Cancer Res* 1996;56:2488–92.
5. Gollin SM. Chromosomal alterations in squamous cell carcinomas of the head and neck: window to the biology of disease. *Head Neck* 2001;23:238–53.
6. Van Houten VM, Tabor MP, van den Brekel MW, Denkers F, Wishaupt RG, Kummer JA, et al. Molecular assays for the diagnosis of minimal residual head-and-neck cancer: methods, reliability, pitfalls, and solutions. *Clin Cancer Res* 2000;6: 3803–16.
7. Braakhuis BJM, Tabor MP, Leemans CR, van der Waal I, Snow GB, Brakenhoff RH. Second primary tumors and field cancerization in oral and oropharyngeal cancer: molecular techniques provide new insights and definitions. *Head Neck* 2002;24: 198–206
8. Helmlinger G, Yuan F, Dellian M, Jain RK. Interstitial pH and pO<sub>2</sub> gradients in solid tumors in vivo: high-resolution measurements reveal a lack of correlation. *Nature Med* 1997;3:177–82
9. Reid CB, Snow GB, Brakenhoff RH, Braakhuis BJ. Biologic implications of genetic changes in head and neck squamous cell carcinogenesis. *Aust N Z J Surg* 1997;67:410–6.
10. Forastiere A, Koch W, Trotti A, Sidransky D. Medical progress— head and neck cancer. *N Engl J Med* 2001;345:1890–900
11. Salehinejad J, Sharifi N, Amirchaghmaghi M, Ghazi N, Shakeri MT, Ghazi A. Immunohistochemical expression of p16 protein in oral squamous cell carcinoma and lichen planus. *Ann Diagn Pathol* 2014;18:210–3.
12. Nemes JA, Deli L, Nemes Z, Márton IJ. Expression of p16INK4A, p53, and Rb proteins are independent from the presence of human papillomavirus genes in oral squamous cell carcinoma. *Oral Surg Oral Med Oral Pathol Oral Radiol Endodontology* 2006;102:344–52.
13. Cordon-Cardo C. Mutation of cell cycle regulators. Biological and clinical implications for human neoplasia. *Am J Pathol* 1995;147:545–60
14. Somers KD, Merrick MA, Lopez ME, Incognito LS, Schechter GL, Casey G. Frequent p53 mutations in head and neck cancer. *Cancer Res* 1992;52:5997–6000.
15. Caamano J, Zhang SY, Rosvold EA, Bauer B, Klein-Szanto AJ. p53 alterations in human squamous cell carcinomas and carcinoma cell lines. *Am J Pathol* 1993;142:1131–9.
16. Tjebbes GW, Leppers VD, Straat FG, Tilanus MG, Hordijk GJ, Slootweg PJ. p53 tumor suppressor gene as a clonal marker in head and neck squamous cell carcinoma: p53 mutations in primary tumor and matched lymph node metastases. *Oral Oncol* 1999;35:384–9.
17. Hanken H, Gröbe A, Cachovan G, et al. CCND1 amplification and cyclin D1 immunohistochemical expression in head and neck squamous cell carcinomas. *Clin Oral Investig* 2014;18:269–76.
18. Monteiro LS, Diniz-Freitas M, Warnakulasuriya S, Garcia-Caballero T, Forteza-Vila J, Fraga M. Prognostic significance of cyclins A2, B1, D1, and E1 and CCND1 numerical aberrations in oral squamous cell carcinomas. *Anal Cell Pathol (Amst)* 2018;2018:7253510.
19. Ramos-García P, González-Moles MÁ, González-Ruiz L, et al. Clinicopathological significance of tumor cyclin D1 expression in oral cancer. *Arch Oral Biol* 2019;99:177–82.
20. Ramos-García P, González-Moles MÁ, Ayén Á, et al. Asymmetrical proliferative pattern loss linked to cyclin D1 overexpression in adjacent non-tumour epithelium in oral squamous cell carcinoma. *Arch Oral Biol* 2019;97:12–
21. Ramos-García P, Bravo M, González-Ruiz L, González-Moles M. Significance of cytoplasmic cyclin D1 expression in oral oncogenesis. *Oral Dis* 2018;24:98–102.
22. Ramos-García P, González-Moles MÁ, González-Ruiz L, Ruiz-Ávila I, Ayén Á, Gil-Montoya JA. Prognostic and clinicopathological significance of cyclin D1 expression in oral squamous cell carcinoma: a systematic review and meta-analysis. *Oral Oncol* 2018;83:96–106.
23. Artavanis-Tsakona S, Rand MD, Lake RJ. Notch signaling: cell fate control and signal transduction in development. *Science (80-)* 1999;284:770–776.

24. Weng AP, Adolfo \*, Ferrando A, et al. Activating Mutations of NOTCH1 in Human T Cell Acute Lymphoblastic Leukemia Downloaded from.; 2004.
25. Stransky N, Egloff AM, Tward AD, et al. The Mutational Landscape of Head Squamous Cell Carcinoma. *Science* (80-) 2014;333:1157–1160
26. Cancer Genome Atlas Network T. Comprehensive genomic characterization of head and neck squamous cell carcinomas. *Nature* 2015;517:576–82.
27. Sakamoto K. Notch signaling in oral squamous neoplasia. *Pathol Int* 2016;66:609–17.
28. Pickering CR, Zhang J, Yoo SY, et al. Integrative genomic characterization of oral squamous cell carcinoma identifies frequent somatic drivers. *Cancer Discov* 2013;3:770–81.
29. Song X, Xia R, Li J, et al. Common and complex Notch1 mutations in chinese oral squamous cell carcinoma. *Clin Cancer Res* 2014;20:701–10.
30. Fan H, Paiboonrungruan C, Zhang X, et al. Nrf2 regulates cellular behaviors and Notch signaling in oral squamous cell carcinoma cells. *Biochem Biophys Res Commun* 2017;493:833–9.
31. Seeburg PH, Ullrich A, Mayes EL V, et al. Human epidermal growth factor receptor cDNA sequence and aberrant expression of the amplified gene in A431 epidermoid carcinoma cells. *Nature* 2004;309:418–25.
32. Lo HW, Hung MC. Nuclear EGFR signalling network in cancers: Linking EGFR pathway to cell cycle progression, nitric oxide pathway and patient survival. *Br J Cancer* 2006;94:184–8.
33. Kalyankrishna S, Grandis JR. Epidermal growth factor receptor biology in head and neck cancer. *J Clin Oncol* 2006;24:2666–72.
34. Grandis JR, Melhem MF, Barnes EL, Tweardy DJ. Quantitative immunohistochemical analysis of transforming growth factor-  $\alpha$  and epidermal growth factor receptor in patients with squamous cell carcinoma of the head and neck. *Cancer* 1996;78:1284–92.
35. Chen IH, Chang JT, Liao CT, Wang HM, Hsieh LL, Cheng AJ. Prognostic significance of EGFR and Her-2 in oral cavity cancer in betel quid prevalent area. *Br J Cancer* 2003;89:681–6.
36. Cully M, You H, Levine AJ, Mak TW. Beyond PTEN mutations: The PI3K pathway as an integrator of multiple inputs during tumorigenesis. *Nat Rev Cancer* 2006;6:184–92.
37. Matsuo FS, Andrade MF, Loyola AM, et al. Pathologic significance of AKT, mTOR, and GSK3 $\beta$  proteins in oral squamous cell carcinoma-affected patients. *Virchows Arch* 2018;472:983–97.
38. Martins F, de Sousa SCOM, dos Santos E, Bin Woo S, Gallottini M. PI3K–AKT–mTOR pathway proteins are differently expressed in oral carcinogenesis. *J Oral Pathol Med* 2016;45:746–52.
39. Zhang H, Liu J, Fu X, Yang A. Identification of key genes and pathways in tongue squamous cell carcinoma using bioinformatics analysis. *Med Sci Monit* 2017;23:5924–32
40. Wang H, Deng X, Zhang J, et al. Elevated expression of zinc finger protein 703 promotes cell proliferation and metastasis through PI3K/AKT/GSK-3 $\beta$  signalling in oral squamous cell carcinoma. *Cell Physiol Biochem* 2017;44:920–34.
41. Yang H, Wen L, Wen M, et al. FoxM1 promotes epithelial-mesenchymal transition, invasion, and migration of tongue squamous cell carcinoma cells through a c-met/ akt-dependent positive feedback loop. *Anticancer Drugs* 2018;29:216–26.
42. Zhang H, Sun JD, Yan L, Jian, Zhao XP. PDGF-D/PDGFR $\beta$  promotes tongue squamous carcinoma cell (TSCC) progression via activating p38/AKT/ERK/EMT signal pathway. *Biochem Biophys Res Commun* 2016;478:845–51.
43. Ito K, Ota A, Ono T, et al. Inhibition of Nox1 induces apoptosis by attenuating the AKT signaling pathway in oral squamous cell carcinoma cell lines. *Oncol Rep* 2016;36:2991–8.
44. Zhang X, Liu N, Ma D, et al. Receptor for activated C kinase 1 (RACK1) promotes the progression of OSCC via the AKT/mTOR pathway. *Int J Oncol* 2016;49:539–48.
45. Sager R. Expression genetics in cancer: shifting the focus from DNA to RNA. *Proc Natl Acad Sci USA* 1997;94:952–5.
46. Helbing CC, Veillette C, Riabowol K, Johnston RN, Garkavtsev I. A novel candidate tumor suppressor, ING1, is involved in the regulation of apoptosis. *Cancer Res* 1997;57:1255–8.
47. Garkavtsev I, Riabowol K. Extension of the replicative life span of human diploid fibroblasts by inhibition of the p33ING1 candidate tumor suppressor. *Mol Cell Biol* 1997;17:2014–9.
48. Garkavtsev I, Kazarov A, Gudkov A, Riabowol K. Suppression of the novel growth inhibitor p33ING1

- promotes neoplastic transformation. *Nat Genet* 1996;14:415—20.
49. Yu GZ, Zhu MH, Zhu Z, Ni CR, Zheng JM, Li FM. Genetic alterations and reduced expression of tumor suppressor p33(ING1b) in human exocrine pancreatic carcinoma. *World J Gastroenterol* 2004;10:3597—601
50. Zeremski M, Horrigan SK, Grigorian IA, Westbrook CA, Gudkov AV. Localization of the candidate tumor suppressor gene ING1 to human chromosome 13q34. *Somat Cell Mol Genet* 1997;23: 233—6.
51. Bignell GR, Warren W, Seal S, Takahashi M, Rapley E, Barfoot R et al. Identification of the familial cylindromatosis tumoursuppressor gene. *Nat Genet* 2000;25:160—5.
52. Biggs PJ, Wooster R, Ford D, Chapman P, Mangion J, Quirk Y, et al. Familial cylindromatosis (turban tumour syndrome) gene localised to chromosome 16q12—q13: evidence for its role as a tumour suppressor gene. *Nat Genet* 1995;11:441—3.
53. Brummelkamp TR, Nijman SM, Dirac AM, Bernards R. Loss of the cylindromatosis tumour suppressor inhibits apoptosis by activating NF-kappaB. *Nature* 2003;424:797—801.
54. Biggs PJ, Chapman P, Lakhani SR, Burn J, Stratton MR. The cylindromatosis gene (cyld1) on chromosome 16q may be the only tumour suppressor gene involved in the development of cylindromas. *Oncogene* 1996;12:1375—7
55. Nybrone O, Bock E. Structure and function of the neural cell adhesion molecules NCAM and L1. *Adv Exp Med Biol* 1990;265:185—96.
56. Barthels D, Vopper G, Boned A, Cremer H, Wille W. High degree of NCAM diversity generated by alternative RNA splicing in brain and muscle. *Eur J Neurosci* 1992;4:327—37.
57. Einheber S, Hannocks M-J, Metz CN, Rifkin DB, Salzer JL. Transforming growth factor-b1 regulates Axon/Schwann cell interactions. *J Cell Biol* 1995;129:443—58.
58. Seki H, Koyama K, Tanaka J, Sato Y, Umezawa A. Neural cell adhesion molecule and perineural invasion in gallbladder cancer. *J Surg Oncol* 1995;58:97—100.
59. Seki H, Tanaka J, Sato Y, Kato Y, Umezawa A, Koyama K. Neural cell adhesion molecule (NCAM) and perineural invasion in bile duct cancer. *J Surg Oncol* 1993;53:78—83.
60. Gandour-Edwards R, Kapadia SB, Barnes L, Donald PJ, Janecka IP. Neural cell adhesion molecule in adenoid cystic carcinoma invading the skull base. *Otolaryngol Head Neck Surg* 1997;117: 453—8.
61. Nakashima M, Sonoda K, Watanabe T. Inhibition of cell growth and induction of apoptotic cell death by the human tumor associated antigen RCAS1. *Nat Med* 1999;5:938—42.
62. Sonoda K, Nakashima M, Saito T, Amada S, Kamura T, Nakano H, et al. Establishment of a new human uterine cervical adenocarcinoma cell line, SiSo, and its reactivity to anti-cancer reagents. *Int J Oncol* 1995;6:1099—104
63. Fukuda M, Tanaka A, Hamao A, Suzuki S, Kusama K, Sakashita H. Expression of RCAS1 and its function in human squamous cell carcinoma of the oral cavity. *Oncol Rep* 2004;12: 259—67.
64. Wiley SR, Schooley K, Smolak PJ, Din WS, Huang CP, Nicholl JK, et al. Identification and characterization of a new member of the TNF family that induces apoptosis. *Immunity* 1995;3: 673—82.
65. Pitti RM, Marsters SA, Ruppert S, Donahue CJ, Moore A, Ashkenazi A. Induction of apoptosis by Apo-2 ligand, a new member of the tumor necrosis factor cytokine family. *J Biol Chem* 1996;271:12687—90
66. Degli-Esposti MA, Smolak PJ, Walczak H, Waugh J, Huang CP, DuBose RF, et al. Cloning and characterization of TRAIL-R3, a novel member of the emerging TRAIL receptor family. *J Exp Med* 1997;186:1165—70.
67. Sheridan JP, Marsters SA, Pitti RM, Gurney A, Skubatch M, Baldwin D, et al. Control of TRAIL-induced apoptosis by a family of signaling and decoy receptors. *Science* 1997;277: 818—21
68. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001;357:539—45.
69. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860—7.
70. Zha S, Yegnasubramanian V, Nelson WG, Isaacs WB, De Marzo AM. Cyclooxygenases in cancer: progress and perspective. *Cancer Lett* 2004;215:1—20.
71. Langowski JL, Zhang X, Wu L, Mattson JD, Chen T, Smith K, et al. IL-23 promotes tumour incidence and growth. *Nature* 2006;27:461—5.
72. Brunda MJ, Luistro L, Warriar RR, Wright RB, Hubbard BR, Murphy M, et al. Antitumor and antimetastatic activity of interleukin 12 against murine tumors. *J Exp Med* 1993;178: 1223—30.

73. Nastala CL, Edington HD, McKinney TG, Tahara H, Nalesnik MA, Brunda MJ, et al. Recombinant IL-12 administration induces tumor regression in association with IFN-production. *J Immunol* 1994;153:1697—706.
74. Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, et al. Divergent pro- and anti-inflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. *J Exp Med* 2003;198:1951—7.
75. Karin M, Cao Y, Greten FR, Li ZW. NF- $\kappa$ B in cancer: from innocent bystander to major culprit. *Nat Rev Cancer* 2002;2:301—10.
76. Orłowski RZ, Baldwin Jr AS. NF- $\kappa$ B as a therapeutic target in cancer. *Trends Mol Med* 2002;8:385—9.
77. Li Q, Verma IM. NF- $\kappa$ B regulation in the immune system. *Nat Rev Immunol* 2002;2:725—34.
78. Maxwell PH, Dachs GU, Gleadle JM, Nicholls LG, Harris AL, Stratford IJ, et al. Hypoxia-inducible factor-1 modulates gene expression in solid tumors and influences both angiogenesis and tumor growth. *Proc Natl Acad Sci USA* 1997;94:8104—9.
79. Ryan HE, Lo J, Johnson RS. HIF-1 $\alpha$  is required for solid tumor formation and embryonic vascularization. *EMBO J* 1998;17:3005—15.
80. Stroka DM, Burkhardt T, Desbaillets I, Wenger RH, Neil DA, Bauer C, et al. HIF-1 is expressed in normoxic tissue and displays an organ-specific regulation under systemic hypoxia. *FASEB J* 2001;15:2445—53.
81. Akakura N, Kobayashi M, Horiuchi I, Suzuki A, Wang J, Chen J, et al. Constitutive expression of hypoxia-inducible factor-1 $\alpha$  renders pancreatic cancer cells resistant to apoptosis induced by hypoxia and nutrient deprivation. *Cancer Res* 2001;61(17):6548—54.
82. Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M, et al. Role of HIF-1 $\alpha$  in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature* 1998;394:485—90.
83. Feinberg AP, Vogelstein B. Hypomethylation of ras oncogenes in primary human cancers. *Biochem Biophys Res Commun* 1983;111:47—54.
84. Andrew PF, Bert V. Hypomethylation distinguishes genes of some human cancers from their normal counterparts. *Nature* 1983;301:89—92.
85. Allameh A, Moazeni-Roodi A, Harirchi I, et al. Promoter DNA methylation and mRNA expression level of p16 gene in oral squamous cell carcinoma: correlation with clinicopathological characteristics. *Pathol Oncol Res* 2018.
86. Hasegawa M, Nelson HH, Peters E, Ringstrom E, Posner M, Kelsey KT. Patterns of gene promoter methylation in squamous cell cancer of the head and neck. *Oncogene* 2002;21:4231—6.
87. Strzelczyk JK, Krakowczyk L, Owczarek AJ. Aberrant DNA methylation of the p16, APC, MGMT, TIMP3 and CDH1 gene promoters in tumours and the surgical margins of patients with oral cavity cancer. *J Cancer* 2018;9:1896—904.
88. Lin RK, Hsieh YS, Lin P, et al. The tobacco-specific carcinogen NNK induces DNA methyltransferase 1 accumulation and tumor suppressor gene hypermethylation in mice and lung cancer patients. *J Clin Invest* 2010;120:521—32.
89. Breitling LP, Yang R, Korn B, Burwinkel B, Brenner H. Tobacco-smoking-related differential DNA methylation: 27K discovery and replication. *Am J Hum Genet* 2011;88:450—7.
90. Kordi-Tamandani DM, Ladies MAR, Hashemi M, Moazeni-Roodi A-K, Krishna S, Torkamanzahi A. Analysis of p15 INK4b and p16 INK4a gene methylation in patients with oral squamous cell carcinoma. *Biochem Genet* 2012;50:448—53.
91. Ishida E, Nakamura M, Ikuta M, et al. Promoter hypermethylation of p14ARF is a key alteration for progression of oral squamous cell carcinoma. *Oral Oncol* 2005;41:614—22.
92. Huang KH, Huang SF, Chen IH, Liao CT, Wang HM, Hsieh LL. Methylation of RASSF1A, RASSF2A, and HIN-1 is associated with poor outcome after radiotherapy, but not surgery, in oral squamous cell carcinoma. *Clin Cancer Res* 2009;15:4174—80.
93. Imai T, Toyota M, Suzuki H, et al. Epigenetic inactivation of RASSF2 in oral squamous cell carcinoma. *Cancer Sci* 2008;99:958—66.
94. Almangush A, Heikkinen I, Mäkitie AA, et al. Prognostic biomarkers for oral tongue squamous cell carcinoma: a systematic review and meta-analysis. *Br J Cancer* 2017;117:856—66.
95. Zhao SF, Yang XD, Lu MX, et al. Prognostic significance of VEGF immunohistochemical expression in oral cancer: a meta-analysis of the literature. *Tumor Biol* 2013;34:3165—71.
96. Liu PF, Kang BH, Wu YM, et al. Vimentin is a potential prognostic factor for tongue squamous cell carcinoma among five epithelial-mesenchymal transition-related proteins. *PLoS ONE* 2017;12.

97. Angadi PV, Patil PV, Angadi V, et al. Immunoexpression of epithelial mesenchymal transition proteins E-cadherin,  $\beta$ -catenin, and N-cadherin in oral squamous cell carcinoma. *Int J Surg Pathol* 2016;24:696–703.
98. Bu J-Q, Chen F. TGF-beta1 promotes cells invasion and migration by inducing epithelial mesenchymal transformation in oral squamous cell carcinoma. *Eur Rev Med Pharmacol Sci* 2017;21:2137–44.
99. Cirillo N, Hassona Y, Celentano A, et al. Cancer-associated fibroblasts regulate keratinocyte cell-cell adhesion via TGF- $\beta$ -dependent pathways in genotype-specific oral cancer. *Carcinogenesis* 2017;38:76–85.
100. Laxmidevi LB, Angadi PV, Pillai RK, Chandreshekar C. Aberrant  $\beta$ -catenin expression in the histologic differentiation of oral squamous cell carcinoma and verrucous carcinoma: an immunohistochemical study. *J Oral Sci* 2010;52:633–40.
101. González-Moles MA, Ruiz-Ávila I, Gil-Montoya JA, Plaza-Campillo J, Scully C.  $\beta$ -Catenin in oral cancer: an update on current knowledge. *Oral Oncol* 2014;50:818–24